



# Comparative Proteomics Kit I: Protein Profiler Module



DNA → RNA → PROTEIN → TRAIT

# Protein Profiler Kit

## Instructors



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# Is There Something Fishy About Teaching Evolution?

**Explore Biochemical Evidence for Evolution**

## Why Teach Protein Electrophoresis?



- **Powerful teaching tool**
- **Real-world connections**
- **Laboratory extensions**
- **Tangible results**
- **Link to careers and industry**
- **Standards-based**

### Scientific Inquiry

- Use of gel electrophoresis to fingerprint proteins
- Use of experimental controls
- Creation of standard curves and cladograms
- Interpretation of experimental results

### Chemistry of Life

- Chemical and physical properties of proteins
- Protein structure (1°, 2°, 3°, 4°) and function
- Protein extraction techniques
- Chemistry of protein electrophoresis

### Genetics

- DNA > RNA > protein > trait
- Molecular vs. morphological classification
- Posttranscriptional and posttranslational modification
- Phylogenetic and cladistic analyses

### Evolution

- Cladistics and phylogenetic relationships
- Biodiversity and natural selection
- Evolution of adaptive traits
- Physical environment as selective force

### Cell and Molecular Biology

- Eukaryotic cell structure and function
- Muscle structure and function

### Environmental and Health Science

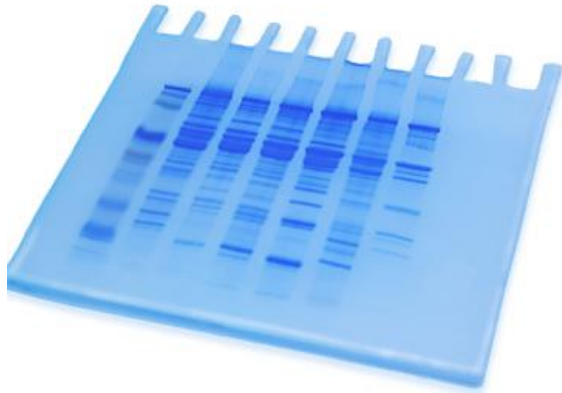
- Interdependence of organisms
- Detecting and analyzing variations in proteins
- Comparative proteomics

## Comparative Proteomics I: Protein Profiler Kit Advantages



- Analyze protein profiles from a variety of fish
- Study protein structure/function
- Use polyacrylamide electrophoresis to separate proteins by size
- Construct cladograms using data from students' gel analysis
- Compare biochemical and phylogenetic relationships. **Hands-on evolution wet lab**
- Sufficient materials for 8 student workstations
- Can be completed in three 45 minute lab sessions

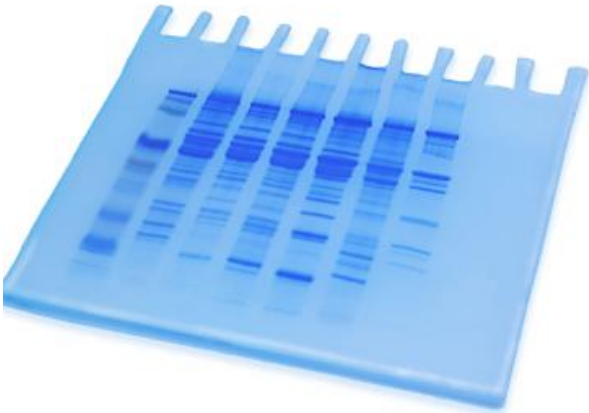
## Workshop Timeline



- **Introduction**
- **Sample Preparation**
- **Load and electrophorese protein samples**
- **Compare protein profiles**
- **Construct cladograms**
- **Stain polyacrylamide gels**
- **Laboratory Extensions**



# Traditional Systematics and Taxonomy



- **Classification**

- Kingdom
- Phylum
- Class
- Order
- Family
- Genus
- Species

- **Traditional classification based upon traits:**

- Morphological
- Behavioral

**Can biomolecular evidence be used to determine evolutionary relationships?**

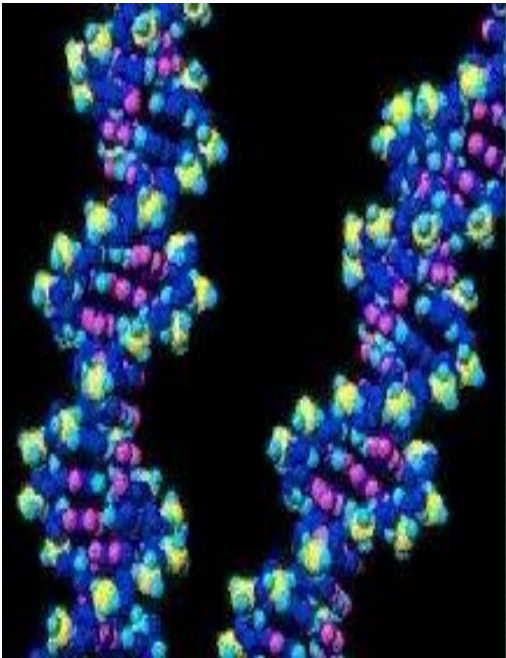
## Biochemical Similarities



- **Traits are the result of:**
  - Structure
  - Function
- **Proteins determine structure and function**
- **DNA codes for proteins that confer traits**

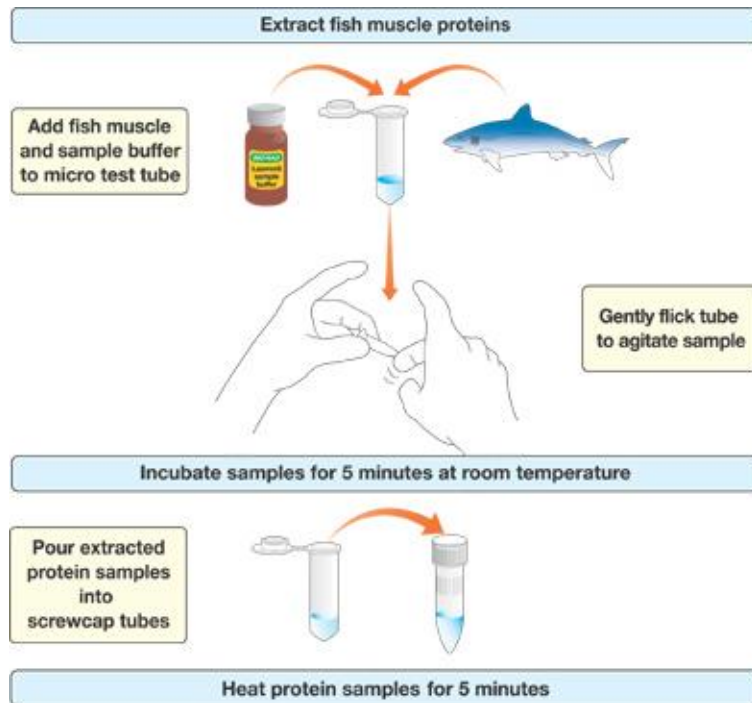
DNA → RNA → PROTEIN → TRAIT

## Biochemical Differences

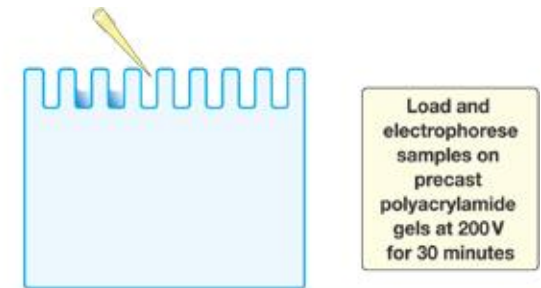


- **Changes in DNA lead to proteins with:**
  - Different functions
  - Novel traits
  - Positive, negative, or no effects
- **Genetic diversity provides pool for natural selection = evolution**

# Protein Fingerprinting Procedures

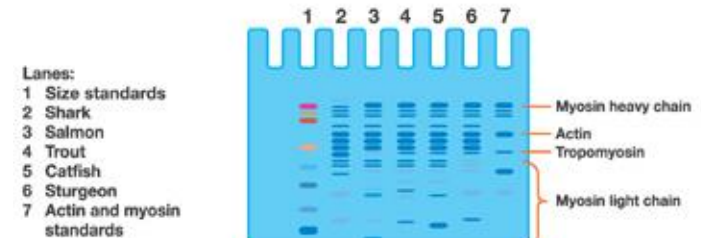


**Day 1**



Stain with Bio-Safe™ Coomassie and destain with water

**Day 2**



Analyze results and dry gels (optional)


Compare molecular data to evolutionary tree

**Day 3**

# Laboratory Quick Guide

## Comparative Proteomics Kit I: Protein Profiler Module – Quick Guide

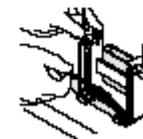
### Lesson 1 Quick Guide

1. Label one 1.5 ml fliplop micro tube for each of five fish samples. Also label one screwcap micro tube for each fish sample.
2. Add 250  $\mu$ l of Bio-Rad Laemmli sample buffer to each labeled fliplop microtube.
3. Cut a piece of each fish muscle about 0.25 x 0.25 x 0.25 cm<sup>3</sup> (  ) and transfer each piece into a labeled fliplop micro test tube. Close the lids.
4. Flick the microtubes 15 times to agitate the tissue in the sample buffer.
5. Incubate for 5 minutes at room temperature.
6. Carefully transfer the buffer by pouring from each fliplop microtube into a labeled screwcap microtube. Do not transfer the fish!
7. Heat the fish samples in screwcap microtubes for 5 minutes at 95°C.



### Lesson 2 Quick Guide

1. Set up MINI-PROTEAN 3 gel box and add 1x TGS electrophoresis buffer to the chamber.
2. Prepare a Ready Gel cassette by cutting along the black line on the bottom of the cassette with a razor blade and pulling off the plastic strip, as indicated on gel cassette.
3. Remove the comb from the Ready Gel cassette.
4. Place Ready Gel cassette into the electrode assembly with the short plate facing inward. Place a buffer dam or another Ready Gel cassette on the opposite side of the electrode assembly, with notch on buffer dam facing inward.



QUICK GUIDE

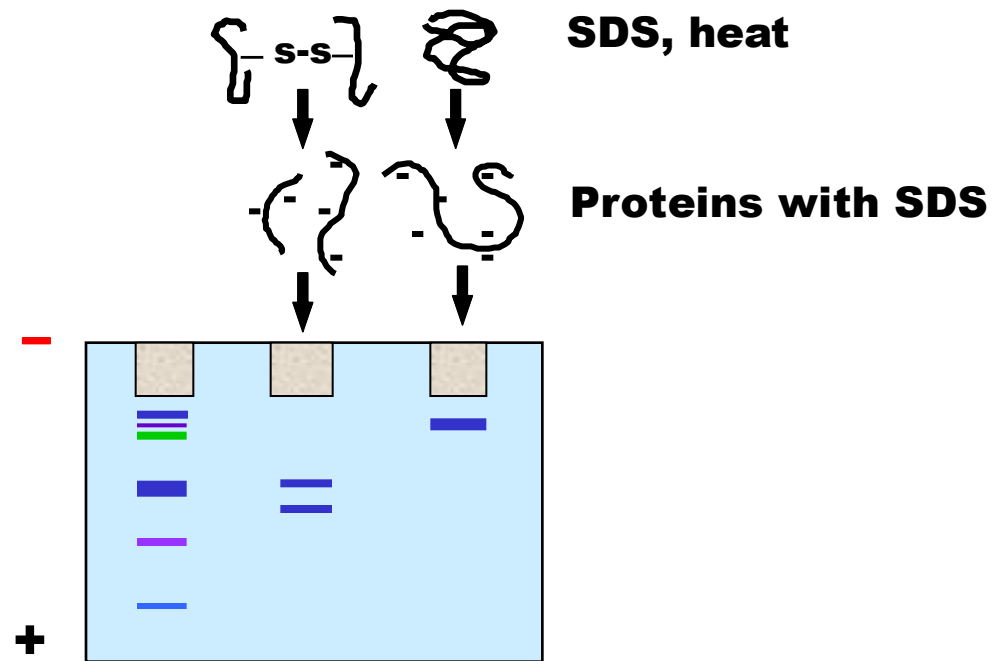
## What's in the Sample Buffer?



- **Tris** buffer to provide appropriate pH
- **SDS** (sodium dodecyl sulfate) detergent to dissolve proteins and give them a negative charge
- **Glycerol** to make samples sink into wells
- **Bromophenol Blue** dye to visualize samples

## Why Heat the Samples?

- **Heating** the samples **denatures** protein complexes, allowing the separation of individual proteins by size





## Making Proteins

**DNA**

TAC    GGA    TCG    AGA    TGA

**mRNA**

AUG    CCU    AGC    UCU    ACU

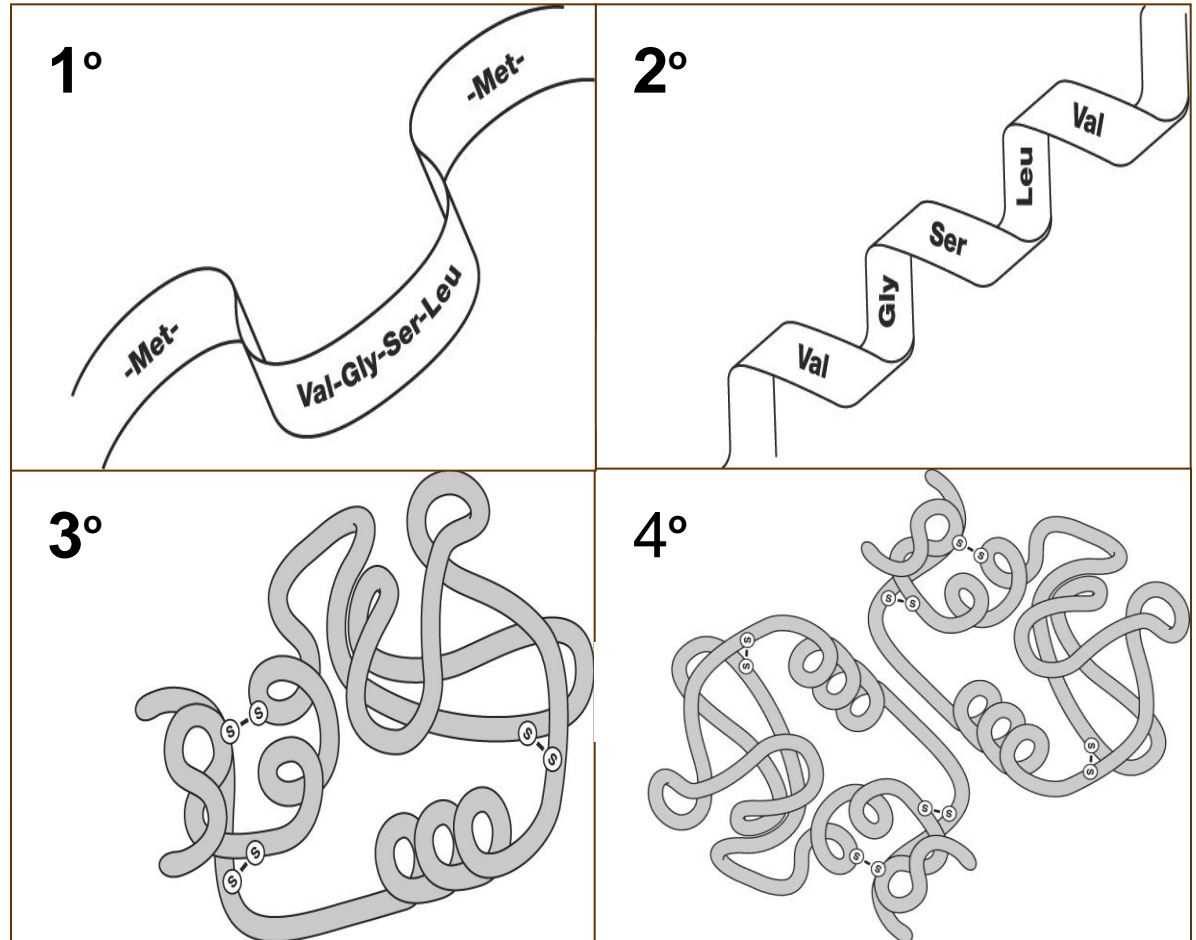
**tRNA**

UAC    GGA    UCG    AGA    UGA

**Amino Acid**

Tyr    Gly    Ser    Arg    STOP

# Levels of Protein Organization

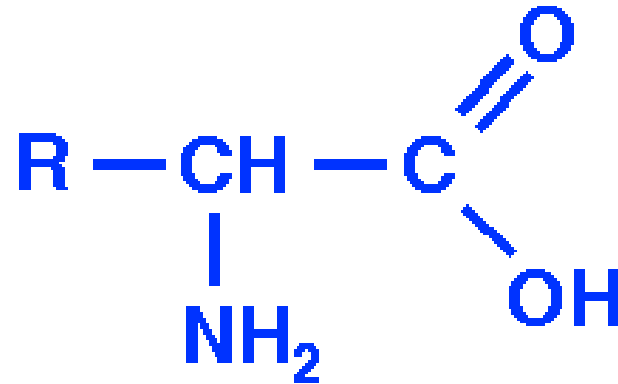


## Protein Size Comparison

- **Break protein complexes into individual proteins**
- **Denature proteins using detergent and heat**
- **Separate proteins based on size**

## Protein Size

- **Size measured in kilodaltons (kD)**
- **Dalton = approximately the mass of one hydrogen atom or  $1.66 \times 10^{-24}$  gram**
- **Average amino acid = 110 daltons**



# Muscle Contains Proteins of Many Sizes

Protein	kD	Function
Titin	3000	Center myosin in sarcomere
Dystrophin	400	Anchoring to plasma membrane
Filamin	270	Cross-link filaments
<b>Myosin heavy chain</b>	<b>210</b>	<b>Slide filaments</b>
Spectrin	265	Attach filaments to plasma membrane
Nebulin	107	Regulate actin assembly
$\alpha$ -actinin	100	Bundle filaments
Gelsolin	90	Fragment filaments
Fimbrin	68	Bundle filaments
<b>Actin</b>	<b>42</b>	<b>Form filaments</b>
Tropomyosin	35	Strengthen filaments
<b>Myosin light chain</b>	<b>15-25</b>	<b>Slide filaments</b>
Troponin (T.I.C.)	30, 19, 17	Mediate contraction
Thymosin	5	Sequester actin monomers

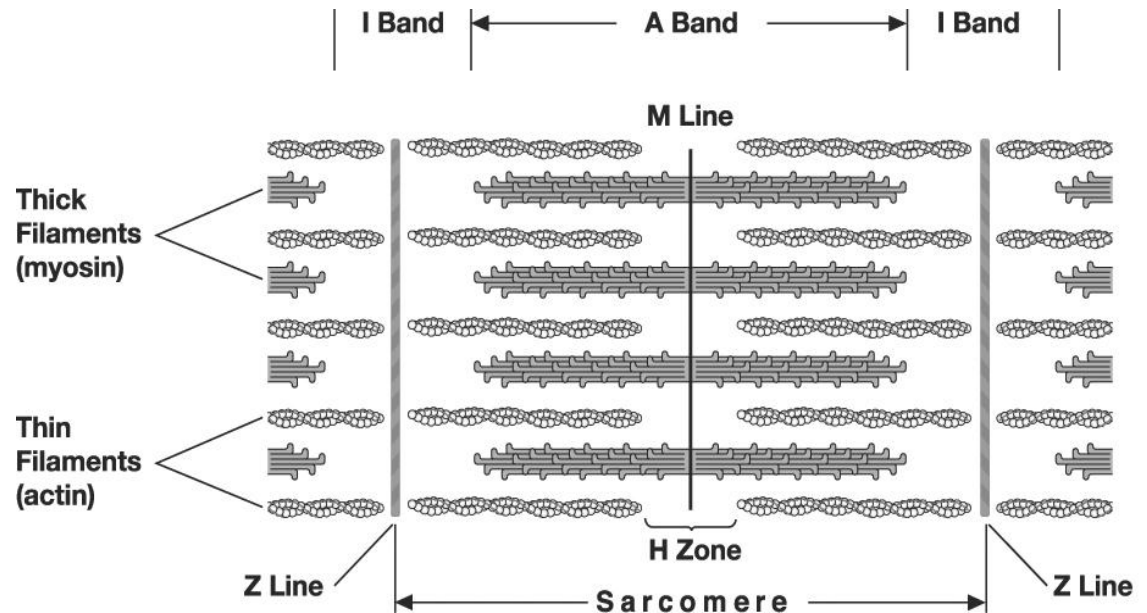
# Actin and Myosin

- **Actin**

- 5% of total protein
- 20% of vertebrate muscle mass
- 375 amino acids = 42 kD
- Forms filaments

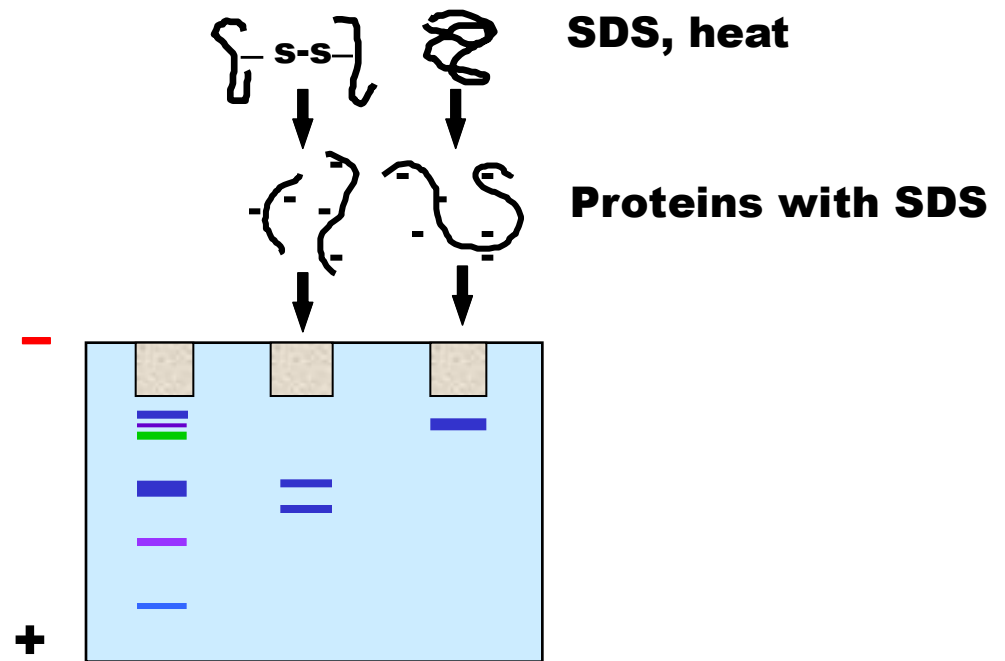
- **Myosin**

- Tetramer
- two heavy subunits (220 kD)
- two light subunits (15-25 kD)
- Breaks down ATP for muscle contraction



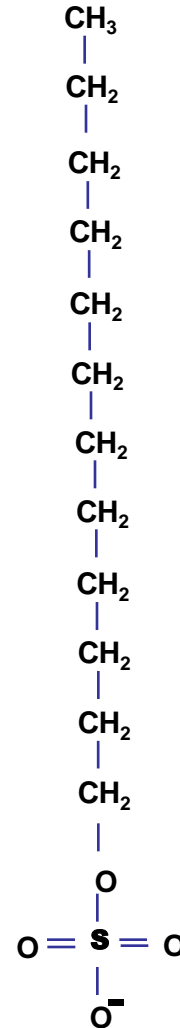
## How Does an SDS-PAGE Gel Work?

- **Negatively charged proteins** move to positive electrode
- **Smaller proteins** move faster
- **Proteins** separate by size



# SDS- Polyacrylamide Gel Electrophoresis (SDS-PAGE)

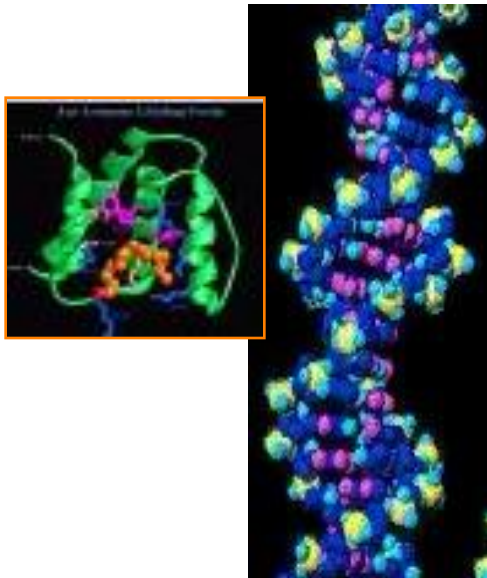
- **SDS detergent**  
(sodium dodecyl sulfate)
  - Solubilizes and denatures proteins
  - Adds negative charge to proteins
- **Heat denatures proteins**



SDS

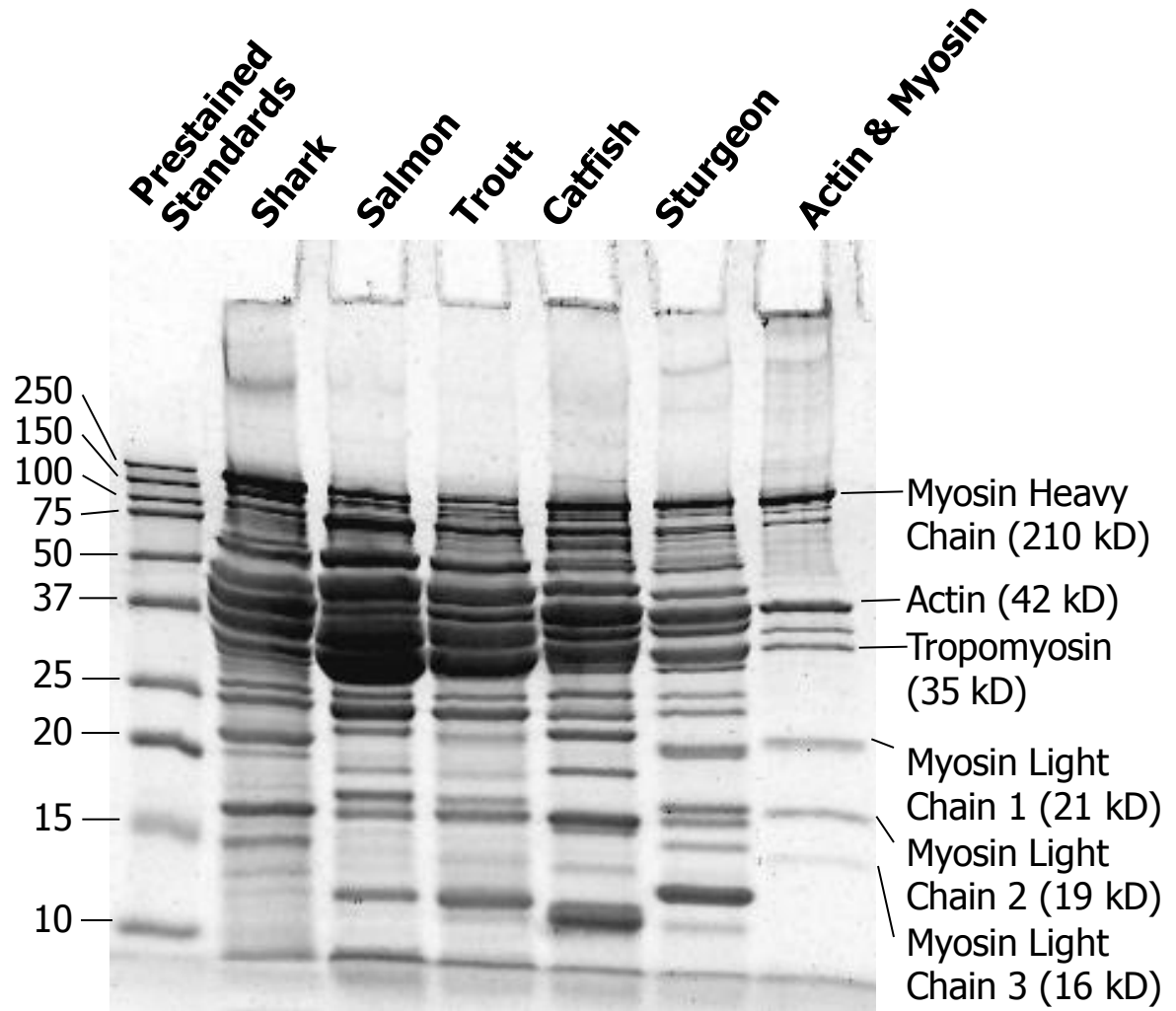


## Why Use Polyacrylamide Gels to Separate Proteins?

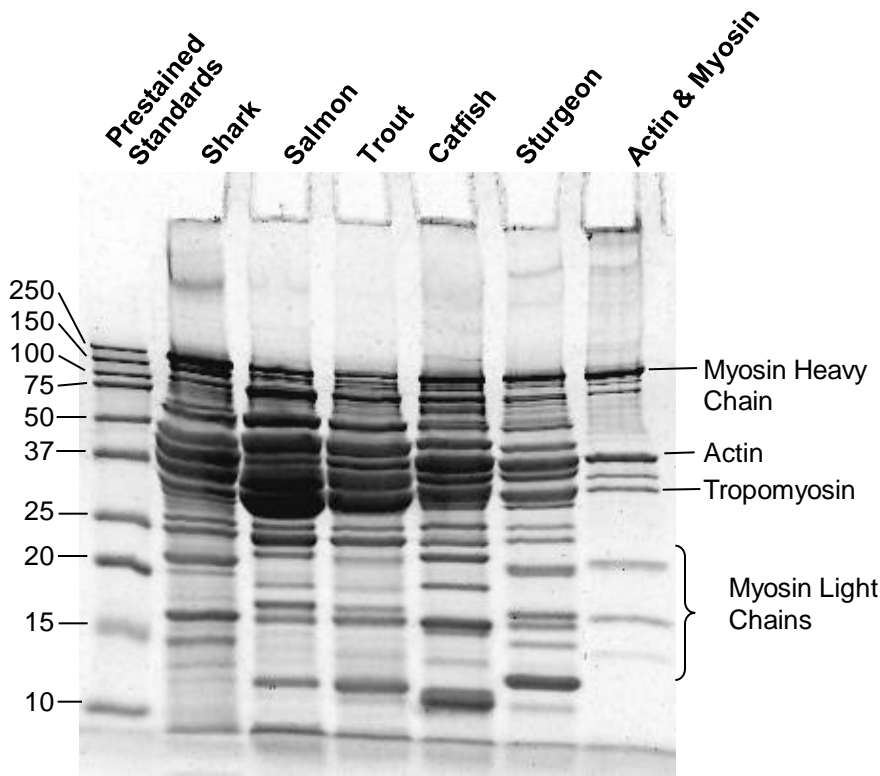


- **Polyacrylamide gel has a tight matrix**
- **Ideal for protein separation**
- **Smaller pore size than agarose**
- **Proteins much smaller than DNA**
  - Average amino acid = 110 daltons
  - Average nucleotide pair = 649 daltons
  - 1 kilobase of DNA = 650 kD
  - 1 kilobase of DNA encodes 333 amino acids = 36 kD

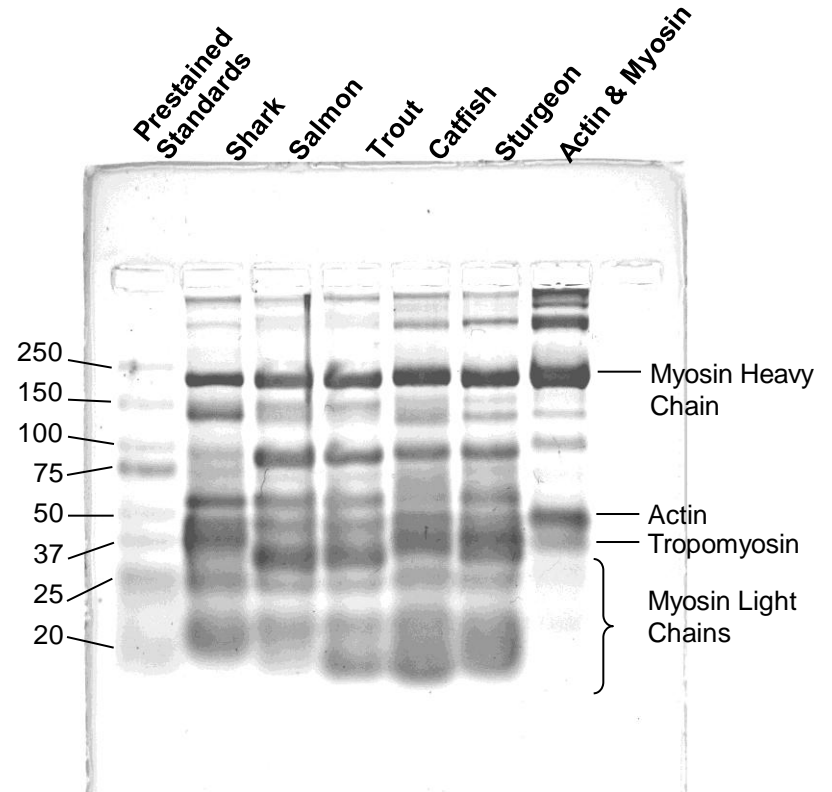
# Polyacrylamide Gel Analysis



# Can Proteins be Separated on Agarose Gels?



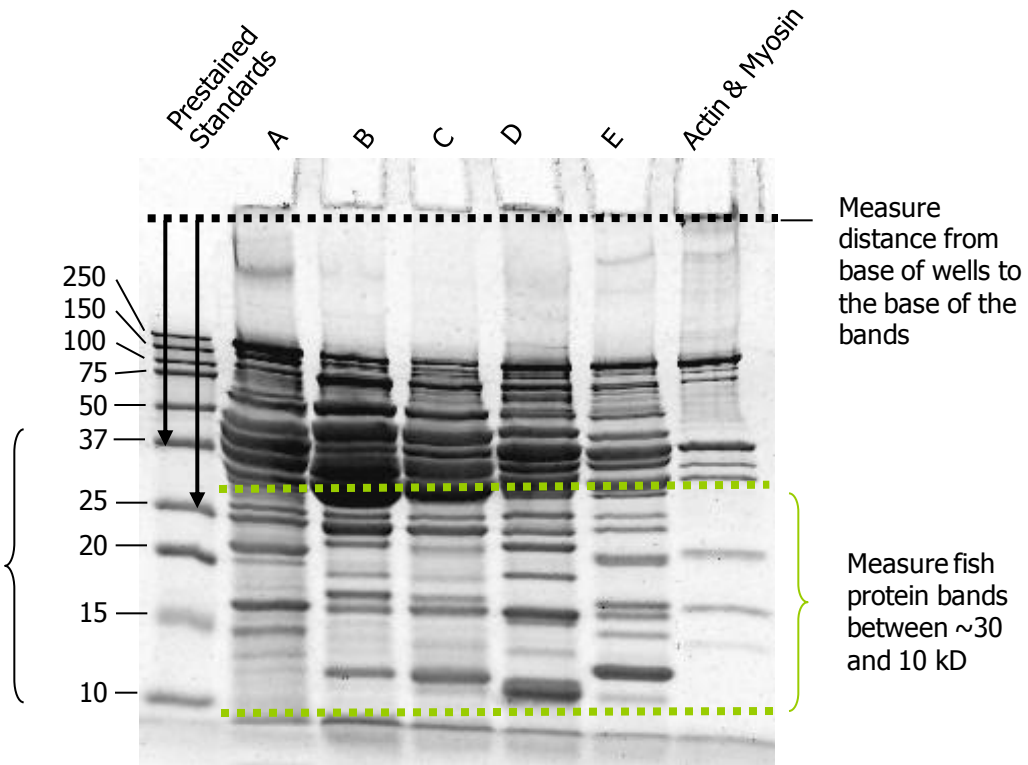
Polyacrylamide



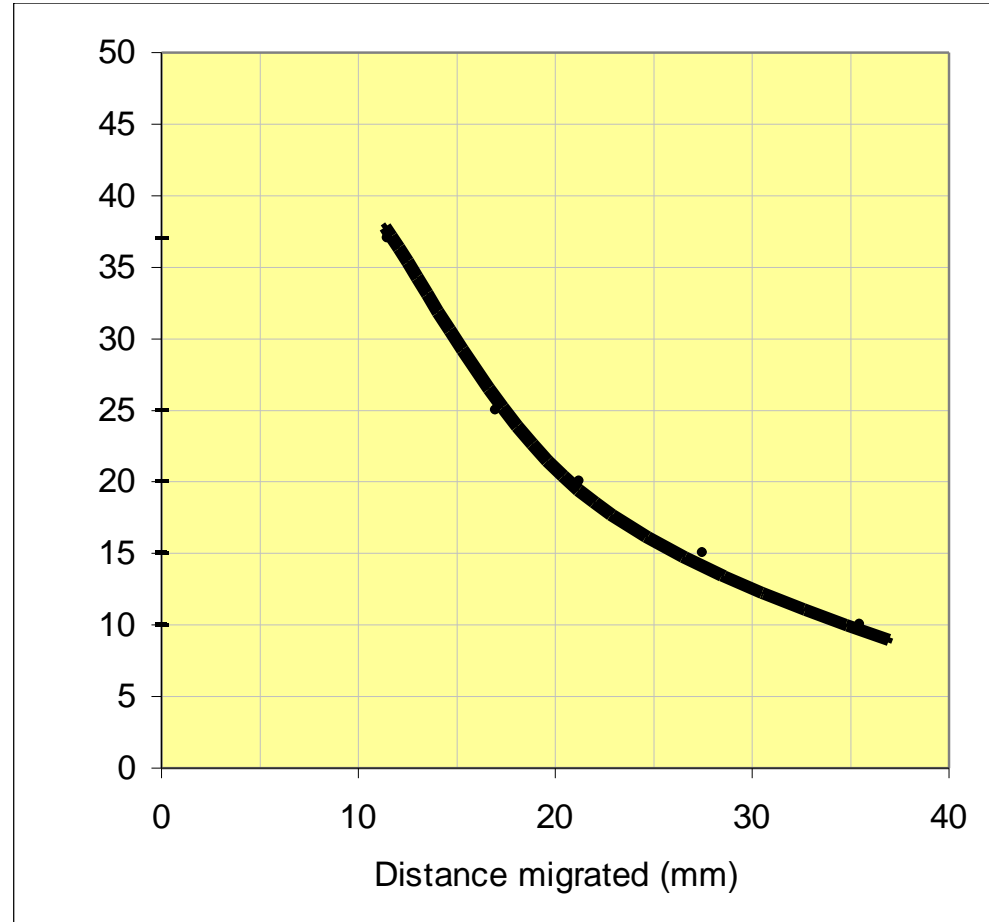
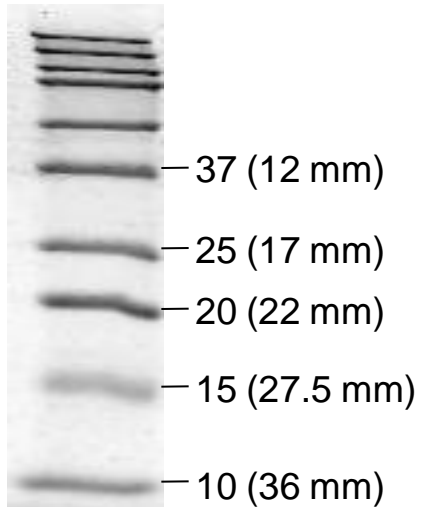
Agarose

# Determine Size of Fish Proteins

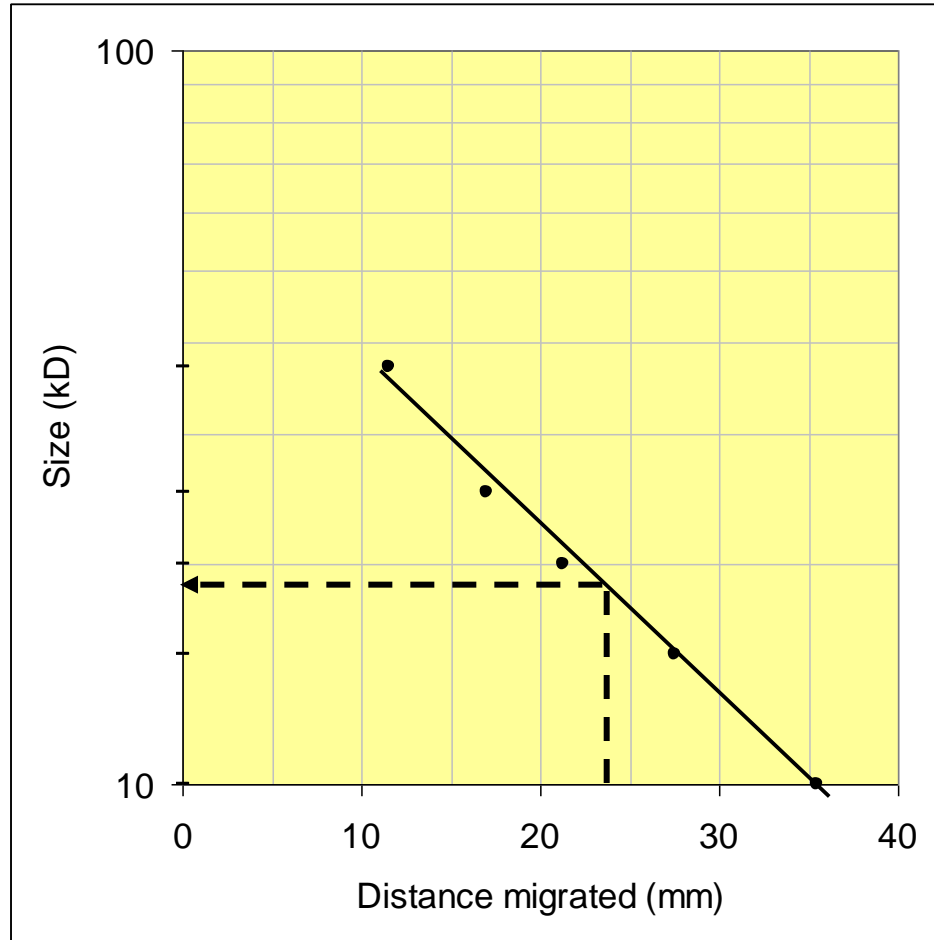
Measure prestained standard bands between ~30 and 10 kD



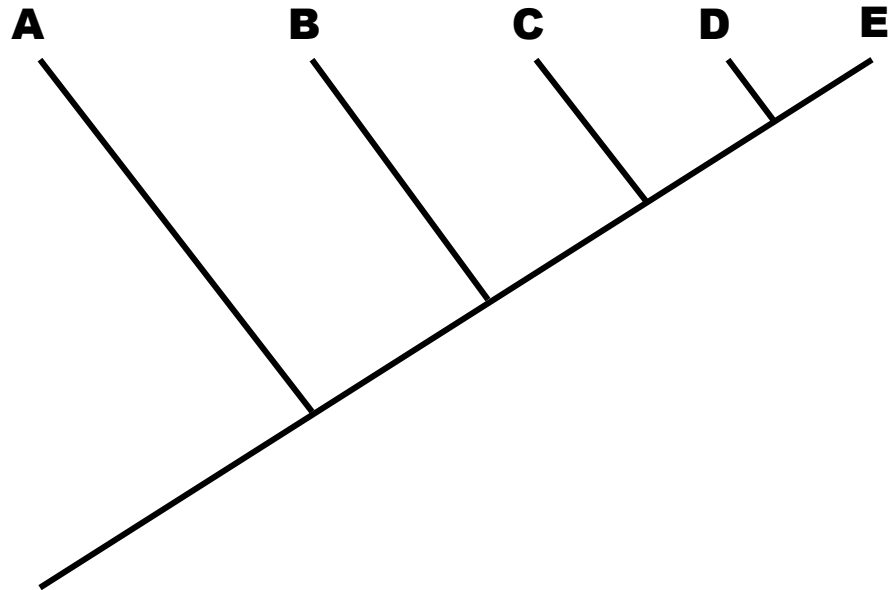
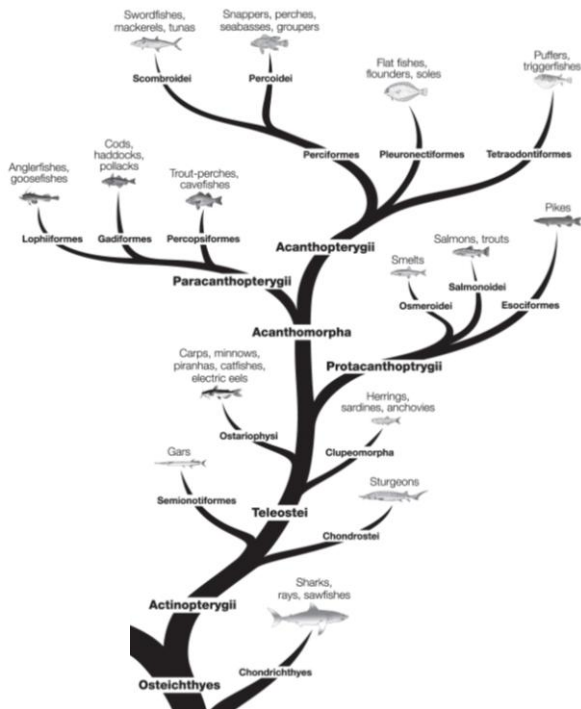
# Molecular Mass Estimation



# Molecular Mass Analysis With Semi-log Graph Paper



# Using Gel Data to Construct a Phylogenetic Tree or Cladogram



# Each Fish Has a Distinct Set of Proteins

	<b>Shark</b>	<b>Salmon</b>	<b>Trout</b>	<b>Catfish</b>	<b>Sturgeon</b>
<b>Total # proteins</b>	<b>8</b>	<b>10</b>	<b>13</b>	<b>10</b>	<b>12</b>
<b>Distance proteins migrated (mm)</b>	25, 26.5, 29, 36, 36.5, 39, 44, 52	26, 27.5, 29, 32, 34.5, 36.5, 37.5, 40.5, 42, 45	26, 27.5, 29, 29.5, 32, 34.5, 36.5, 37.5, 40.5, 42, 45, 46.5, 51.5	26, 27.5, 29, 32, 36.5, 38, 38.5, 41, 46, 47.5	26, 27.5, 30, 30.5, 33, 35.5, 37, 39, 39.5, 42, 44, 47

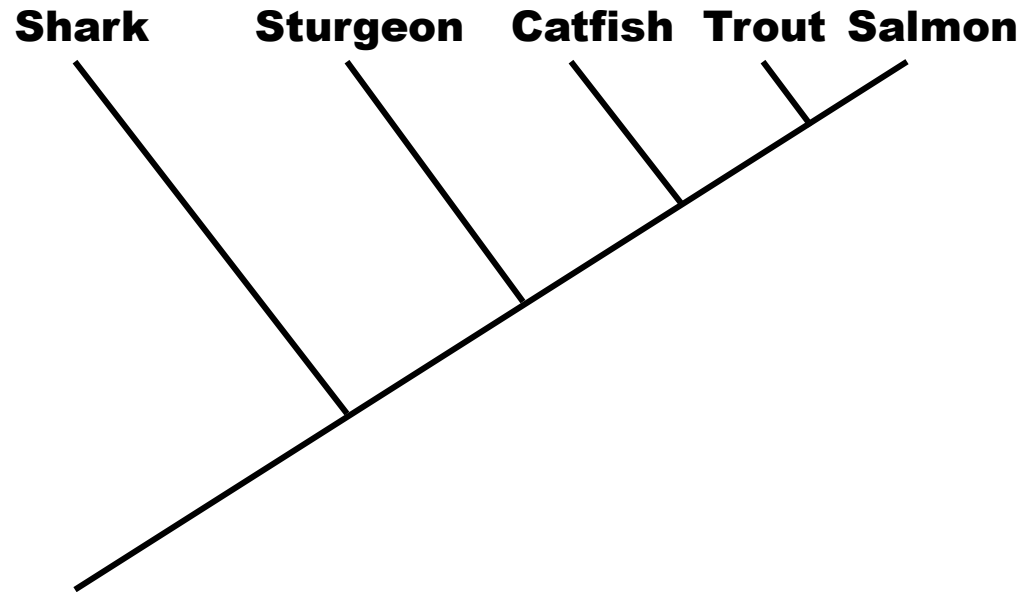


# Some of Those Proteins Are Shared Between Fish

Distance (mm)	Size (kD)	Shark	Salmon	Trout	Catfish	Sturgeon
25	32.5	X				
26	31.5		X	X	X	X
26.5	31.0	X				
27.5	30.0		X	X	X	X
28.5	29.1					
29	28.6	X	X	X	X	
30	27.6			X		X
30.5	27.1					X
32	25.6		X	X	X	
33	24.7					X
34.5	23.2		X	X		
35.5	22.2					X
36	21.7	X				
36.5	21.2	X	X	X	X	
37	20.7					X
37.5	20.2		X	X		
38	19.7				X	
38.5	19.3				X	

# Character Matrix Is Generated and Cladogram Constructed

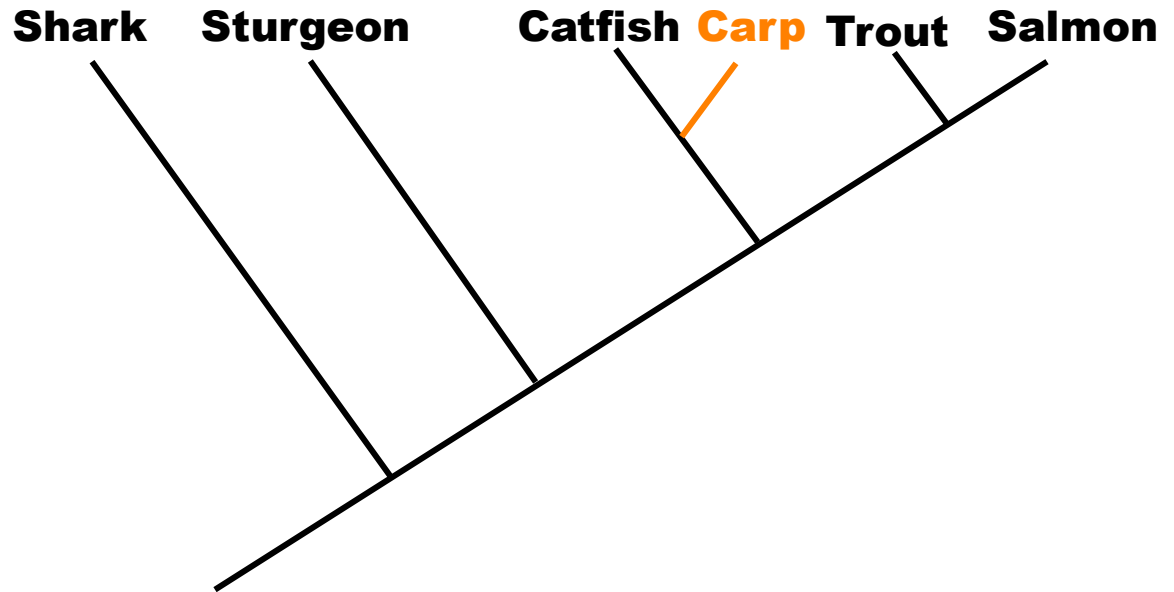
	<b>Shark</b>	<b>Salmon</b>	<b>Trout</b>	<b>Catfish</b>	<b>Sturgeon</b>
<b>Shark</b>	8	2	2	2	2
<b>Salmon</b>	2	10	10	5	3
<b>Trout</b>	2	10	13	5	4
<b>Catfish</b>	2	5	5	10	2
<b>Sturgeon</b>	2	3	4	2	12





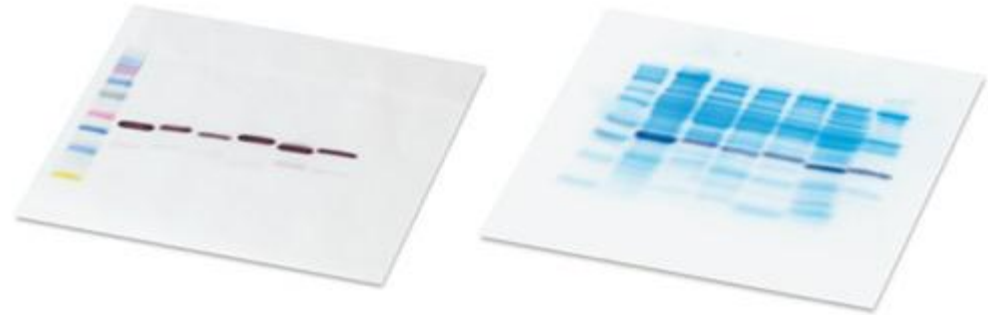
# Pairs of Fish May Have More in Common Than to the Others

	Shark	Salmon	Trout	Catfish	Sturgeon	Carp
Shark	8	2	2	2	2	2
Salmon	2	10	10	5	3	5
Trout	2	10	13	5	4	5
Catfish	2	5	5	10	2	8
Sturgeon	2	3	4	2	12	2
Carp	2	5	5	8	2	11

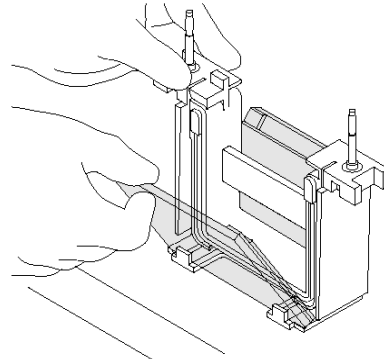
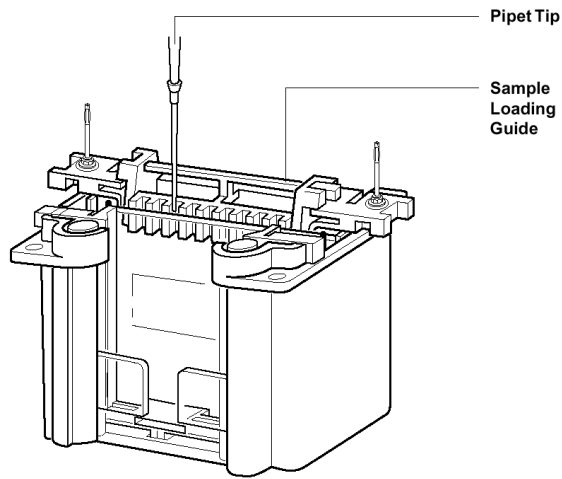


## Extensions

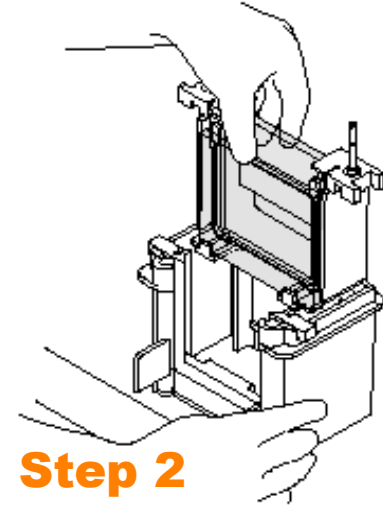
- **Independent study**
- **Western blot analysis**



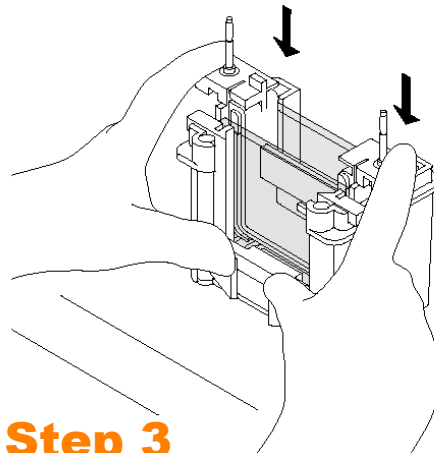
# Ready Gel® Precast Gel Assembly



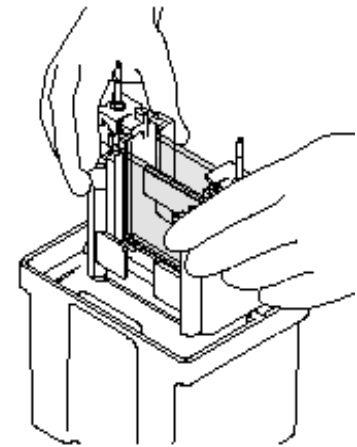
**Step 1**



**Step 2**



**Step 3**



**Step 4**